

LIGNANS FROM *MACHILUS THUNBERGII*

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Key Word Index—*Machilus thunbergii*; Lauraceae; 2,5-diaryl-3,4-dimethyltetrahydrofuran type neolignans; machilin-F, G, H, I; (−)-nectandrin-A.

Abstract—Five new neolignans, machilin-F [2-(4-hydroxy-3-methoxyphenyl)-3,4-dimethyl-5-piperonyltetrahydrofuran], machilin-G [2-(3,4-dimethoxyphenyl)-3,4-dimethyl-5-piperonyltetrahydrofuran], machilin-H [2-(4-hydroxy-3,5-dimethoxyphenyl)-3,4-dimethyl-5-(4-hydroxy-3-methoxyphenyl)tetrahydrofuran], machilin-I [2,5-bis-(4-hydroxy-3-methoxyphenyl)-3,4-dimethyltetrahydrofuran], (−)-nectandrin-A and the known compound, nectandrin-B were isolated from the bark of *Machilus thunbergii* and their structure were determined.

INTRODUCTION

In a previous paper [1], we reported the isolation of the neolignans, machilin-A–E, from the bark of *Machilus thunbergii* Sieb. et Zucc. The methanol extract of the bark of this plant was further investigated and 2,5-diaryl-3,4-dimethyltetrahydrofuran type neolignans, machilin-F–I, were isolated we now report their structures elucidated by means of spectral analysis.

RESULTS AND DISCUSSION

Machilin-F (**1**) was obtained as a colourless oil. In the EI-mass spectrum, a molecular ion peak was observed at *m/z* 342. The ¹H NMR and IR spectra indicated that **1** was a diastereomer of austrobailignan-7 [2] and chicanine [3] belonging to the 2,5-diaryl-3,4-dimethyltetrahydrofuran type of neolignans.

The chemical shifts of H-2 (δ4.44), H-5 (4.46) and 3,4-Me (1.01) in the ¹H NMR spectrum indicated each methyl group to be situated in a *trans*-relation to the vicinal aryl group [4, 5]. The configuration of **1** was thus concluded to be either the all *trans* form or the *meso* form and the chemical shifts of H-3 and H-4 (δ2.19) indicated the 3,4-*cis*-relationship [6]. Furthermore, compound **1** showed no Cotton effect (200–350 nm), but known compounds of the all *trans*-configuration did so in circular dichroism [7]. Thus, the configuration for the structure of **1** was found to be the *meso*-configuration with the 3-Me and 4-Me groups situated in a *cis*-relationship to each other. From these data, the structure of **1** is thus shown to be (2*R*^{*,} 3*R*^{*,} 4*S*^{*,} 5*R*^{*)}-2-(4-hydroxy-3-methoxyphenyl)-3,4-dimethyl-5-piperonyltetrahydrofuran.

Compounds **2–5** were each obtained as a colourless oil. From the ¹H and ¹³C NMR spectral data, these compounds may also possibly be derivatives of the 2,5-diaryl-3,4-dimethyltetrahydrofuran type of neolignan. Their relative configurations were shown to be the same as that of **1** by the ¹H NMR, specific rotation and circular dichroism data.

Machilin-G (**2**) showed a molecular ion peak at *m/z* 356 in the EI-mass spectrum, and its IR spectrum showed no absorption band for a hydroxyl group. The above data

and ¹H NMR spectrum indicated **2** to be (2*R*^{*,} 3*R*^{*,} 4*S*^{*,} 5*R*^{*)}-2-(3,4-dimethoxyphenyl)-3,4-dimethyl-5-piperonyltetrahydrofuran, a diastereomer of calopiptin [6, 8].

Compounds **3** and **4** were identified as nectandrin-B and an enantiomer of nectandrin-A, respectively, by comparison with data in the literature [9].

Machilin-H (**5**) showed absorption band(s) of hydroxyl group(s) in its IR spectrum. A molecular ion peak was noted at *m/z* 374 in the EI-mass spectrum. The ¹H NMR and ¹³C NMR spectra indicated the structure of **5** to possibly be (2*R*^{*,} 3*R*^{*,} 4*S*^{*,} 5*R*^{*)}-2-(4-hydroxy-3,5-dimethoxyphenyl)-3,4-dimethyl-5-(4-hydroxy-3-methoxyphenyl)tetrahydrofuran.

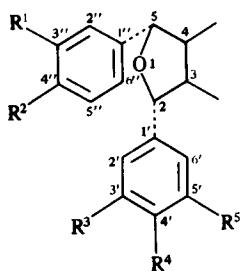
Machilin-I (**6**) was obtained as a colourless oil. From the IR, ¹H NMR and EI-mass spectral data, it was concluded to be a diastereomer of nectandrin-B. The signals of methine (δ5.45) and methyl groups (0.61) due to H-2 and 3-Me, respectively, of this compound shifted further downfield and upfield than those of **1** by the anisotropic effect of the aromatic group at C-2 [4, 5]. The methyl group at C-3 thus appears to be in a *cis*-relationship to the aromatic group at C-2. By measuring the NOE-correlated 2D NMR spectrum of **6**, the *cis*-configuration between the methyl groups at C-3 and C-4 was established. The relative configuration of this compound may thus be represented as (2*S*^{*,} 3*R*^{*,} 4*S*^{*,} 5*R*^{*)}-2,5-bis-(4-hydroxy-3-methoxyphenyl)-3,4-dimethyltetrahydrofuran.

Compounds **1**, **2**, **4–6** have not been reported previously as naturally occurring lignans.

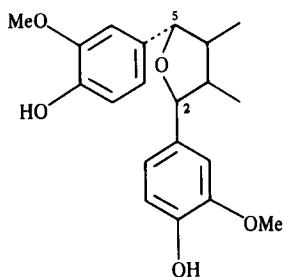
EXPERIMENTAL

NMR spectra were measured at 400 MHz for ¹H NMR and 100 MHz for ¹³C NMR. Chemical shifts were given on the (ppm) scale with TMS as the internal standard. Details of the extraction and isolation are described in the previous paper [1].

Machilin-G (**2**). Colourless oil, 15.3 mg; $[\alpha]_D^{24} 10.8^\circ$ (CHCl₃; *c* 0.40); IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{−1}: 3540, 2930, 1710, 1450, 1230; UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (log *ε*): 282 (3.63), 231 (3.93), 208 (4.24); ¹H NMR (CDCl₃): δ 1.01 (6H, *d*, *J* = 6.7 Hz, 3,4-Me), 2.29 (2H, *m*, H-3,4), 4.44 (1H, *d*, *J* = 6.3 Hz, H-2 or 5), 4.46 (1H, *d*, *J* = 6.7 Hz, H-2 or



	R ¹	R ²	R ³	R ⁴	R ⁵
1	OCH ₂ O	OMe	OH	H	
2	OCH ₂ O	OMe	OMe	H	
3	OMe	OH	OMe	OH	H
4	OMe	OH	OMe	OMe	H
5	OMe	OH	OMe	OH	OMe

**6**

5), 3.90 (3H, s, -OMe), 5.95 (2H, s, OCH₂O), 6.76–6.97 (6H, *m*, aromatic protons); MS (*m/z*): 342 [M]⁺, 190, 175, 145.

Machilin-G (**2**): Colourless oil, 15.3 mg; $[\alpha]_D^{24^\circ}$ 3.9° (CHCl₃; *c* 0.30); IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹: 2960, 1595, 1518, 1263; UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (log *e*): 282 (4.19), 217 (4.45); ¹H NMR (CDCl₃): δ 1.02 (3H, *d*, *J* = 6.7 Hz, 3 or 4-Me), 1.03 (3H, *d*, *J* = 6.7 Hz, 3 or 4-Me), 2.30 (2H, *m*, H-3,4), 3.88 (3H, s, OMe), 3.90 (3H, s, OMe), 4.46 (1H, *d*, *J* = 6.9 Hz, H-2 or 5), 4.47 (1H, *d*, *J* = 6.9 Hz, H-2 or 5), 5.95 (2H, s, OCH₂O), 6.80–6.99 (6H, *m*, Ar-H); MS (*m/z*): 356 [M]⁺, 340, 206, 190, 175, 145.

Nectandrin-B (**3**): Colourless oil, 448 mg; $[\alpha]_D^{23^\circ}$ 0° (CHCl₃; *c* 0.40)

(*-*)-*Nectandrin-A* (**4**): Colourless oil, 80 mg; $[\alpha]_D^{23^\circ}$ -8.9° (CHCl₃; *c* 0.40); IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹: 3550, 2950, 1605, 1250; UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (log *e*): 279 (3.92), 230 (4.30), 210 (4.35); ¹H NMR (CDCl₃): δ 1.05 (3H, *d*, *J* = 6.7 Hz, 3 or 4-Me), 1.04 (3H, *d*, *J* = 6.7 Hz, 3 or 4-Me), 2.30 (2H, *m*, H-3,4), 3.90 (9H, s, OMe), 4.50 (1H, *d*, *J* = 6.2 Hz, H-2 or 5), 4.51 (1H, *d*, *J* = 6.2 Hz, H-2 or 5), 5.55 (1H, *br*, OH), 6.80–7.00 (6H, *m*, Ar-H); MS (*m/z*): 358 [M]⁺, 216, 192, 145.

Machilin-H (**5**): Colourless oil, 7.3 mg; $[\alpha]_D^{25^\circ}$ 8.8° (CHCl₃; *c* 0.37); IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹: 3540, 2930, 1610, 1510, 1460, 1210; UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (log *e*): 279 (3.91), 216 (4.38); ¹H NMR (CDCl₃): δ 1.05 (6H, *d*, *J* = 6.8 Hz, 3, 4-Me), 2.35 (2H, *m*, H-3,4), 3.90 (9H, s, OMe), 4.49 (1H, *d*, *J* = 5.3 Hz, H-2 or 5), 4.50 (1H, *d*, *J* = 5.3 Hz, H-2 or 5), 5.47 (1H, *br*, OH), 5.58 (1H, *br*, OH), 6.65 (2H, *s*, Ar-H), 6.91–6.96 (3H, *m* Ar-H); MS (*m/z*): 374 [M]⁺, 222, 192, 145.

Machilin-I (**6**): Colourless oil, 2.6 mg; $[\alpha]_D^{25^\circ}$ -93.0° (CHCl₃; *c* 0.13); IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹: 3500, 2920, 1500, 1450, 1260, 1200; UV nm (log *e*): 280 (3.85), 210 (4.24); ¹H NMR (CDCl₃): δ 0.61 (3H, *d*, *J* = 7.0 Hz, 3-Me), 1.00 (3H, *d*, *J* = 6.4 Hz, 4-Me), 2.44 (2H, *m*, H-3,4), 3.88 (3H, s, OMe), 3.91 (3H, s, OMe), 4.64 (1H, *d*, *J* = 9.2 Hz, H-5), 5.45 (1H, *d*, *J* = 4.4 Hz, H-2), 5.52 (1H, *br*, OH), 5.56 (1H, *br*, OH), 6.79–6.95 (6H, *m*, Ar-H); MS (*m/z*): 344 [M]⁺, 271, 192, 145, 124.

Table 1. ¹³C NMR chemical shifts (δ) of compounds **1–6**
100 MHz CDCl₃

	1	2	3	4	5	6
1'	134.0	134.7	134.2	134.8	133.1	132.7
2'	107.9	108.0	109.2	109.8	103.2	108.8
3'	146.5	149.0	146.5	148.9	146.9	146.7
4'	145.1	148.5	145.0	148.3	134.0	144.4
5'	114.1	111.1	114.1	111.1	146.9	114.4
6'	119.9	118.6	119.3	118.5	103.2	118.9
2	87.3 ^a	87.4	87.3	87.2 ^a	87.2 ^a	84.8
3	44.4 ^b	44.5	44.3	44.2 ^b	44.5 ^b	47.6
4	44.5 ^b	44.5	44.3	44.3 ^b	44.0 ^b	43.5
5	87.4 ^a	87.4	87.3	87.3 ^a	87.5 ^a	85.8
1''	136.2	136.2	134.2	134.1	134.0	135.1
2''	106.8	106.8	109.2	109.1	109.0	108.4
3''	147.7	147.8	146.5	146.5	146.5	146.3
4''	146.9	147.0	145.0	145.0	145.0	145.1
5''	109.0	109.8	114.1	114.1	114.0	114.1
6''	119.3	119.9	119.3	119.2	120.0	119.3
3-Me	12.8	12.8 ^c	12.9	12.8 ^c	13.2	9.4
4-Me	12.8	12.9 ^a	12.9	12.9 ^c	12.8	11.9
OMe	55.9	56.0	55.8	55.7	55.9	56.0
OMe		55.9	55.8	55.9	56.3	56.0
OMe				55.7	56.3	
OCH ₂ O	100.9	100.9				

^{a,b,c} Assignments are interchangeable.

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CLEOMISCOSIN D, A COUMARINO-LIGNAN FROM SEEDS OF *CLEOME VIScosa*

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Abstract—Cleomiscosin D, a minor coumarino-lignan of the seeds of *Cleome viscosa*, has been proved to be regiosomer of cleomiscosin C. A method of degradation of coumarino-lignans for the identification of the coumarin moiety has been developed.

INTRODUCTION

Cleomiscosins A (1) and B (2), isolated from the seeds of *Cleome viscosa* Linné and fully characterized from comprehensive spectral analysis and some chemical reactions, constitute the first regiosomeric pair of coumarino-lignans in which a coumarin moiety is linked with a phenylpropanoid unit through a dioxane bridge [1-3]. The correctness of these structures were later verified by Merlini and co-workers who achieved their synthesis by oxidative coupling of fraxetin with coniferyl alcohol [4]. We subsequently reported the isolation of a third coumarino-lignan, cleomiscosin C (3) from this source [3] and showed it to be identical with aquillochin [5] for which two alternative structures were proposed. Further investigation on this plant material led us to isolate a known coumarin and a new coumarinolignan, cleomiscosin D. The characterization of these two compounds and a method of cleaving the dioxane bridge of cleomiscosins for identification of coumarin moieties will be discussed in the present paper.

RESULTS AND DISCUSSION

A coumarin, $C_{10}H_8O_5$ (MS m/z 208, M^+), was recognized to be oxygenated at its 6, 7 and 8-positions from

its UV, IR spectra together with its 1H NMR signals for two coumarin hydrogens [δ 6.17 and 7.74 (1H each, d , $J=9.5$ Hz)], a lone aromatic hydrogen (δ 6.61) and a methoxyl grouping (δ 3.88). Irradiation of the methoxyl signal at δ 3.88 showed an observable NOE of the isolated signal at δ 6.61 which in turn showed long range coupling with the coumarin H-4 signal at δ 7.74. The observation clearly revealed it to be fraxetin [4].

Cleomiscosin D (4), $C_{21}H_{20}O_9$ (MS, m/z 416, M^+), responded to tests for phenol. The UV and IR data of cleomiscosin D were essentially identical with those of cleomiscosin C (3), an isomer of cleomiscosin D. The ^{13}C NMR spectrum of cleomiscosin D disclosed the presence of eight aliphatic carbons ($Me-O \times 3$, $-CH_2-O \times 1$, $>CH-O \times 2$, $-CH=CH- \times 1$), 12 aromatic carbons ($CH \times 3$, $C \times 2$, $C-O \times 7$) and one carbonyl-like cleomiscosin C. Cleomiscosin D gave a diacetate (4a), $C_{25}H_{24}O_{11}$ (MS, m/z 500, M^+).

The 1H NMR spectrum of cleomiscosin D in pyridine- d_5 showed signals for three methoxyl groupings (δ 3.77, 3.77 and 3.82), two coumarin hydrogens (δ 6.41 and 7.73), an isolated aromatic hydrogen (δ 6.74) and four aliphatic hydrogens on carbons carrying oxygen functions (δ 3.96, 4.30, 4.57 and 5.55). Significant amounts of NOE were observed between the isolated signal at δ 6.74 and a methoxyl signal at 3.82, between the isolated signal at 6.74